

DCEG Core Genotyping Facility

Stephen Chanock, M.D.

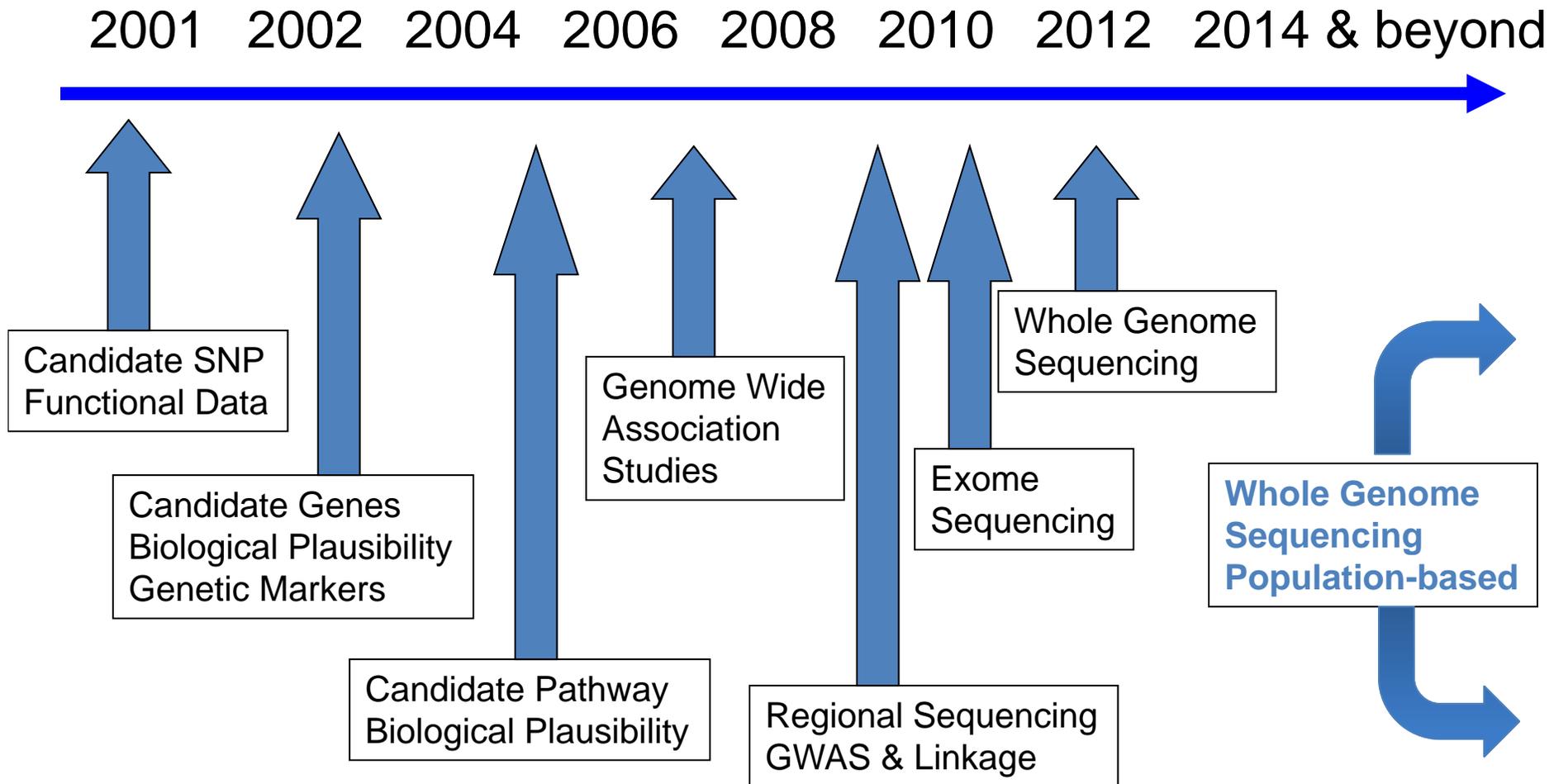
**Chief, Laboratory of Translational Genomics
Director, Core Genotyping Facility**

January 25, 2012

Mission of Core Genotyping Facility (CGF)

- Conduct of high quality molecular epidemiology studies
 - Emphasis on:
 - Germline contribution to risk
 - Gene-environment interactions
 - Transition to:
 - Germline/somatic interactions
 - Interaction of somatic alterations with environmental risk factors
- Education
 - Genetics analysis courses & seminars

Milestones at the Core Genotyping Facility



NATIONAL CANCER INSTITUTE
Division of Cancer Epidemiology and Genetics

Office of the Director
Joseph F. Fraumeni, Jr., M.D.
Director

Administrative Resource Center
Donna Siegle
Director, Office of Administrative Services,
DCEG

**Office of Communications
& Special Initiatives**
Catherine B. McClave, M.S.
Chief

Office of Education
Jackie A. Lavigne, Ph.D., M.P.H.
Chief

**Office of Division
Operations & Analysis**
Marianne K. Henderson, M.S.
Chief

**Epidemiology and
Biostatistics Program**
Robert N. Hoover, M.D., Sc.D.
Director

**Human Genetics
Program**
Margaret A. Tucker, M.D.
Director

Biostatistics Branch
Nilanjan Chatterjee, Ph.D.
Chief

**Hormonal & Reproductive
Epidemiology Branch**
Louise A. Brinton, Ph.D.
Chief

**Genetic
Epidemiology Branch**
Neil E. Caporaso, M.D.
Chief

**Clinical
Genetics Branch**
Mark H. Greene, M.D.
Chief

**Infections & Immuno-
Epidemiology Branch**
Allan Hildesheim, Ph.D.
Chief

**Nutritional
Epidemiology Branch**
Vacant

**NCI Core Genotyping
Facility**
Stephen J. Chanock, M.D.
Director

**Laboratory of
Translational Genomics**
Stephen J. Chanock, M.D.
Chief

**Occupational &
Environmental
Epidemiology Branch**
Debra T. Silverman, Sc.D.
Chief

**Radiation
Epidemiology Branch**
Martha S. Linet, M.D.
Chief

Office of Director of SAIC
Dedicated Support

Core Genotyping Facility (CGF) DNA
Extraction & Sample Handling (DESL)

Basic Research Program
Dedicated Support

Laboratory of Translational Genomics
Genetic Epidemiology Branch Laboratory

**DCEG Activities at the
Frederick Federal Research
and Development Center
(SAIC-F)**

**Applied & Development
Directories (ADD)**

Dedicated Support

Repository Methods
Immunological Monitoring

Shared Services

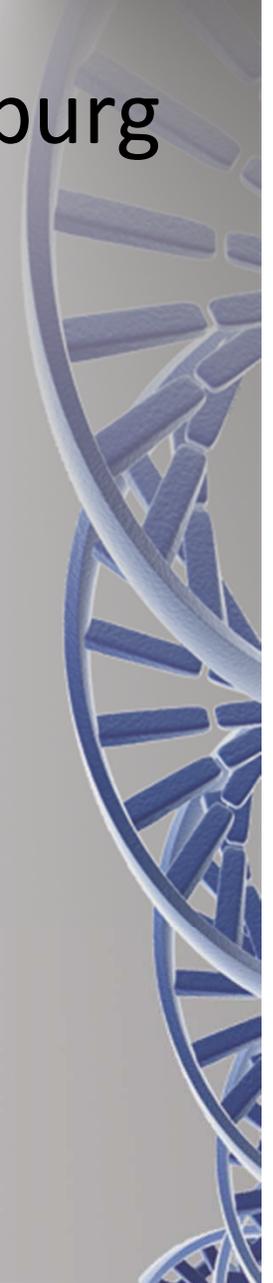
Bioprocessing & Transformations
Repository Support

Advanced Technology Program
Shared Services

Lab of Molecular Technology
Laboratory of Proteomics & Analytical Technology
(LPAT Hormone Unit – dedicated to DCEG)

CGF Facilities Footprint

Advanced Technology Center: Gaithersburg



The Core Genotyping Facility

Dedicated DCEG Facility

What's in a name? Core Plus Plus

Core Services

Genotyping

Sequencing

Computing Support

Data Analysis

+

Collaboration

GWAS & Follow up

Candidate Gene Studies

Regional/ Exome/ Genome Sequencing

Data Sharing

> 500 Publications

+

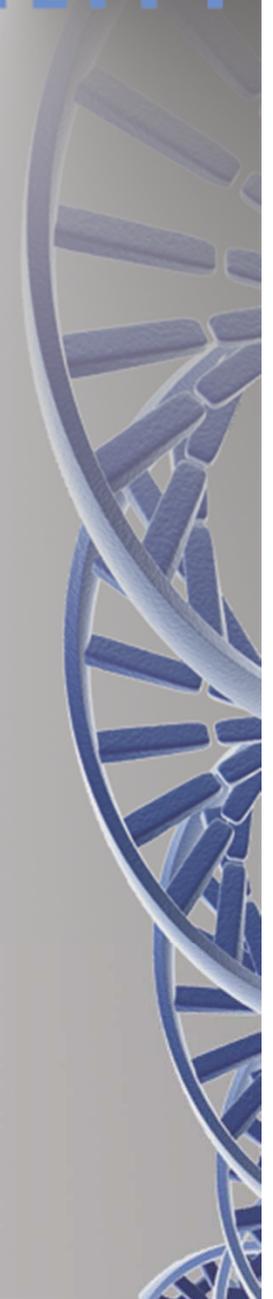
Innovation

Biotechnology

Genomics

Computational Methods

Statistical Methods





Meredith Yeager
Scientific Director

Amy Hutchinson
Operations &
Administration

Open
Production
Laboratory

Joe Boland
Research &
Development

Kevin Jacobs
Bioinformatics &
Analysis



DESL

LIMS

Genotyping

Sequencing

QA/QC

*Technology
Transfer*



Investigation of Alternatives

- DCEG Conducted Molecular Epidemiology Pilot Study 2001-2003
 - 5 Companies asked to produce defined data sets
 - Common issues
 - Slow
 - Costly
 - Poor performance with QC
- Periodic reassessment of contract work
 - Loss of scientific ownership
 - Variability in deliverables

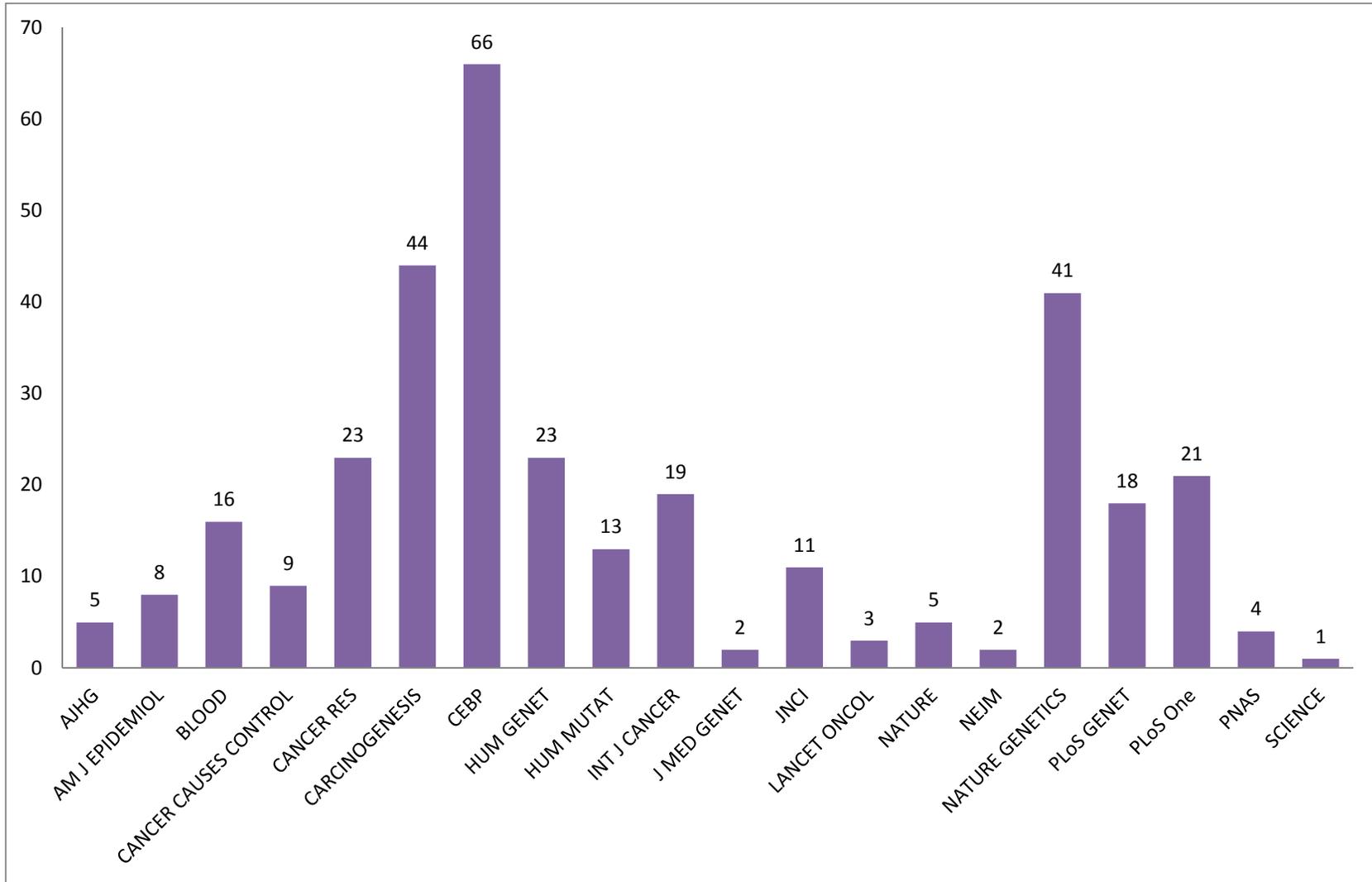
Value of creating CGF within FFRDC

- Close collaboration between NCI investigators and SAIC-F experts
- NCI can monitor every step and assess capacity to meet milestones
- Opportunity to drive scientific challenges in partnership
 - *Bridging Epidemiology and Genetics*

Nimble Personnel Structure

- Reorganization began with 9 SAIC FTEs
 - Reorganization and expansion 2002-2006
 - CGEMS funding for 5 additional analysts
- Current FTEs: 42
 - Shift from wet to dry positions in last 3 years
- Establish expertise for genetic analysis
 - Avoid “blackbox/blackhole” of contract
- Embed NCI oversight within SAIC work flow
 - Daily- no..... hourly discussions

536 CGF Publications for 2002-2011



Review of DCEG Projects for CGF

- Proposals discussed and approved by Branch Chiefs prior to submission
- Varies by scope & cost
 - Senior Leadership for Genomics Committee (SLGC) provides concept review for
 - GWAS chips
 - Sequencing of Exome/Whole Genome
 - Genotype Review Committee (GRC)
 - All projects greater than \$25,000

Senior Leadership for Genomics Committee (SLGC)

Mission

Review & Approval of

GWAS chips

Exome/WGS

Determines priority for

Illumina Infinium

Data Sharing and Access

Issues

Membership

J Fraumeni

P Tucker

R Hoover

P Hartge

S Chanock

M Henderson

Monthly Meetings with Minutes

Genotyping Review Committee (GRC)

Mission

Critique of Science

Statistical Review

Approval letter required
to proceed to CGF
queue

Minutes

Chair can approve small
projects & revisions

Membership

Chair:

P Tucker, Director, HGP

PIs from each Branch

rotate every 2 years

S Chanock

K Pitt

CGF Review Processes

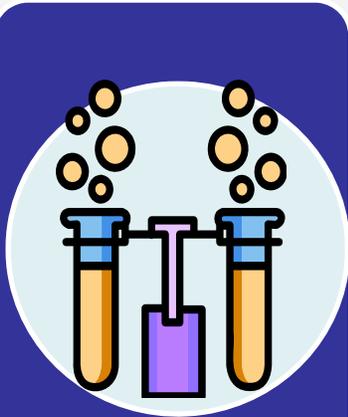
- Weekly conference
- Monthly SLGC meeting
- Quarterly SAIC report
- Biannual review of budget by OD DCEG
- Quadrennial Site Visit
 - May 2012 for CGF

Dedicated Facility Support

- DCEG directly supports
 - Personnel
 - Equipment
 - Maintenance
- Each project competes for DCEG resources

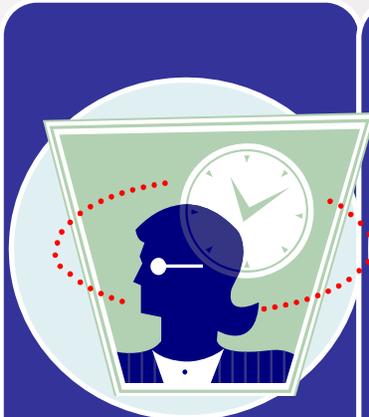
Operations

Research & Development



DESL DNA
Extraction and
Handling

7 staff



Project
Management

3 staff



Administrative
Support

3 staff

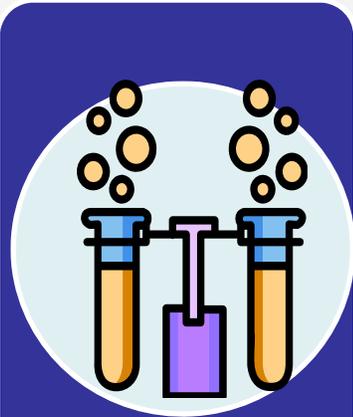


Research and
Development

6 staff

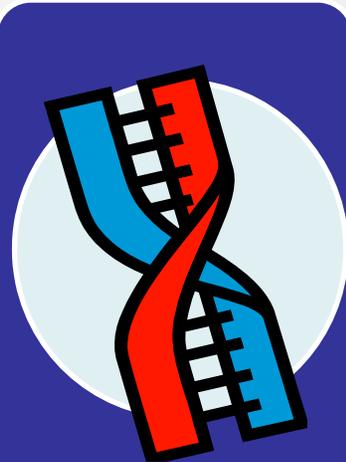


Production Laboratory



Production
Genotyping

6 staff



Production
Sequencing

3 staff



Quality
Assurance &
Control

3 staff



LIMS

4 staff



Technology
Transfer



Director

Critical CGF Laboratory Team



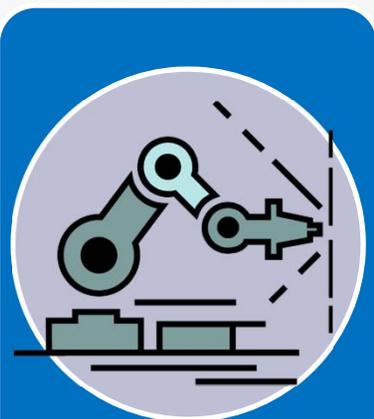
Quality
Assurance &
Control

3 staff

- Review technology performance metrics
- Generate and update:
 - SOPs
 - Staff training
- Equipment maintenance
- Follow-up on laboratory problems
- Cost savings measures



CGF Bioinformatics & Scientific Operations



LIMS,
Database
& Web

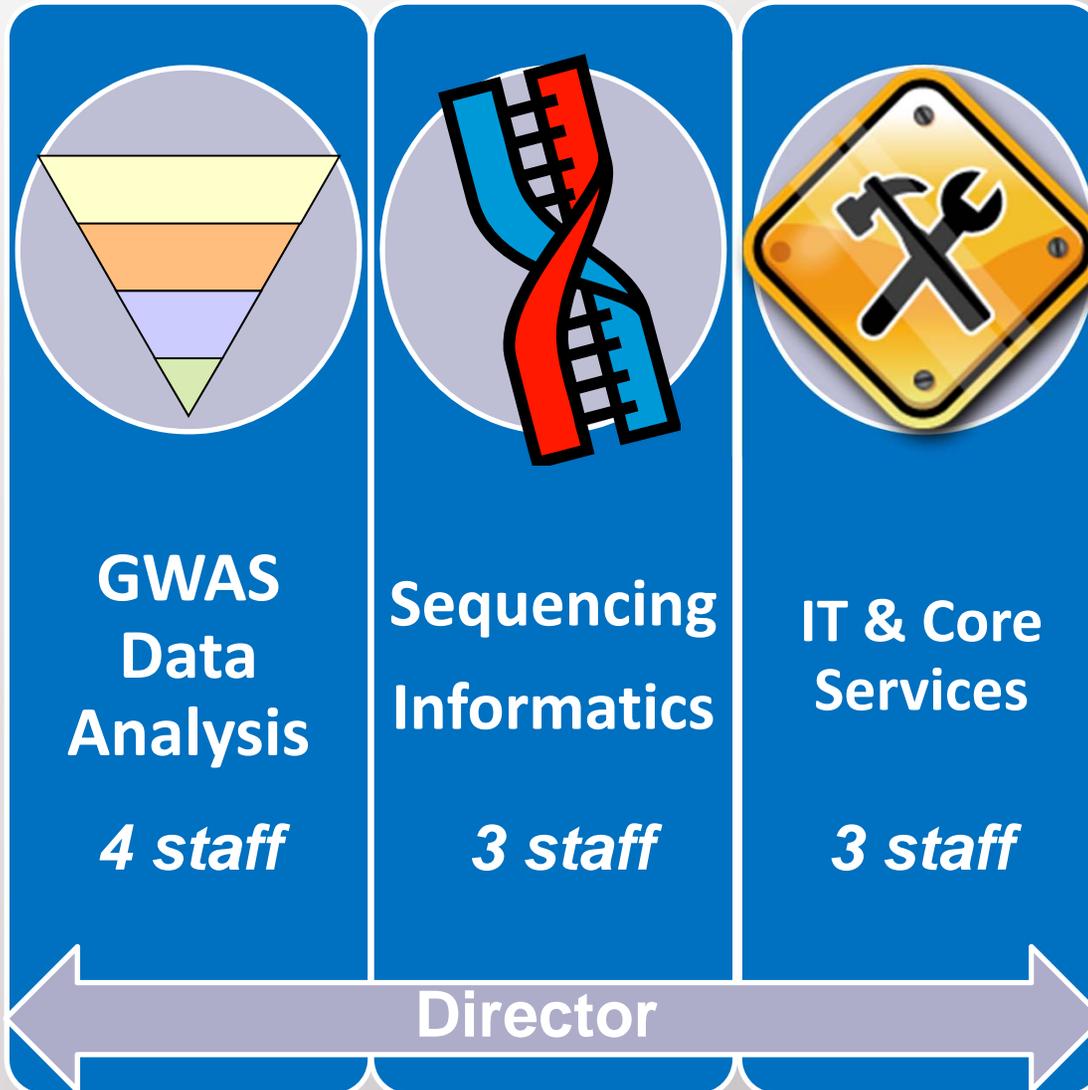
4 staff

- Maintains Commercial LIMS
 - LabVantage 2004
- Customize content for CGF workflow
- Oversees archiving of data
 - Virtual lab note books only
- Oversee security/permissions

- Maintains websites
 - Public CGF
 - <http://cgf.nci.nih.gov/>
 - VariantGPS (replaces SNP500)
 - <http://variantgps.nci.nih.gov>

Bioinformatics & Analysis Version 3.0

Science and informatics at warp speed



Open Source Tools

- GLU software: <http://code.google.com/p/glu-genetics>
- Genotype data
 - SNP array data management
 - Quality control, population structure, & association analysis
- Next-generation sequencing (NGS)
 - Infrastructure to produce and manage alignments
 - Parse and manipulate variants
 - Conversions to/from VCF, GFF, PLINK, BEAGLE, Germline, GLU
 - Annotation of known/novel, function, frequency
 - Efficient *in silico* exome/regional pull-down
 - Visualization tools: Coverage, ploidy, CNV, SV, allelic ratio

Onsite CGF IT Infrastructure



**IT & Core
Services**

3 staff

- High-performance computing clusters
 - Over 640 CPU cores, >2 TB RAM
 - Supporting CGF
 - + DCEG (LTG, BB, REB, GEB)
 - + CCR/SAIC-F Sequencing Facility
- Laboratory instrument support
 - Integrated high performance computing
- Large-scale data storage subsystems
 - Over 300 TB tier 1 storage
- Local and wide-area networking
- Battery and generator backup of computing and HVAC
- Systems administration and security
 - Interface with CBIIT and CIT

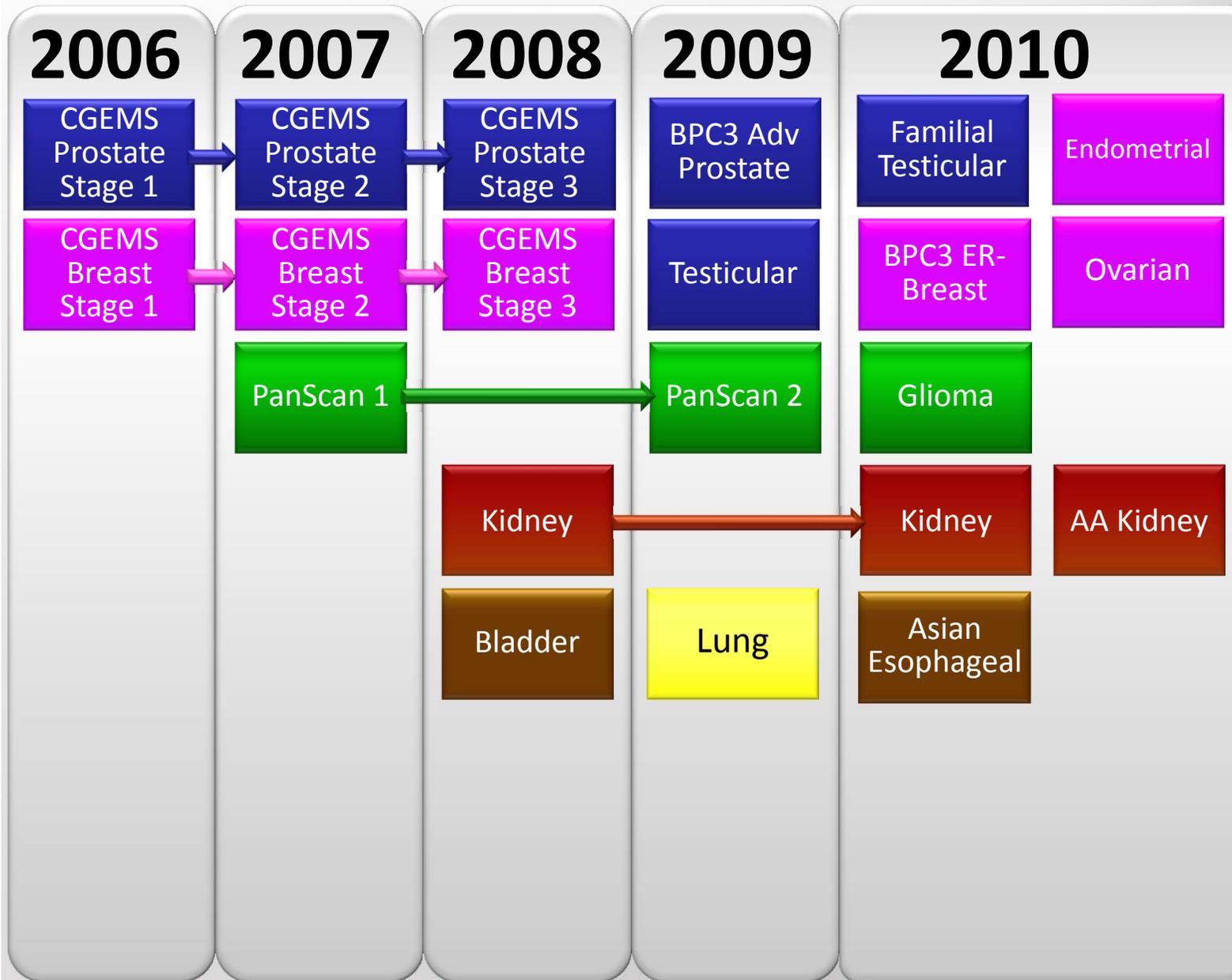


CGF Data Output since 2002

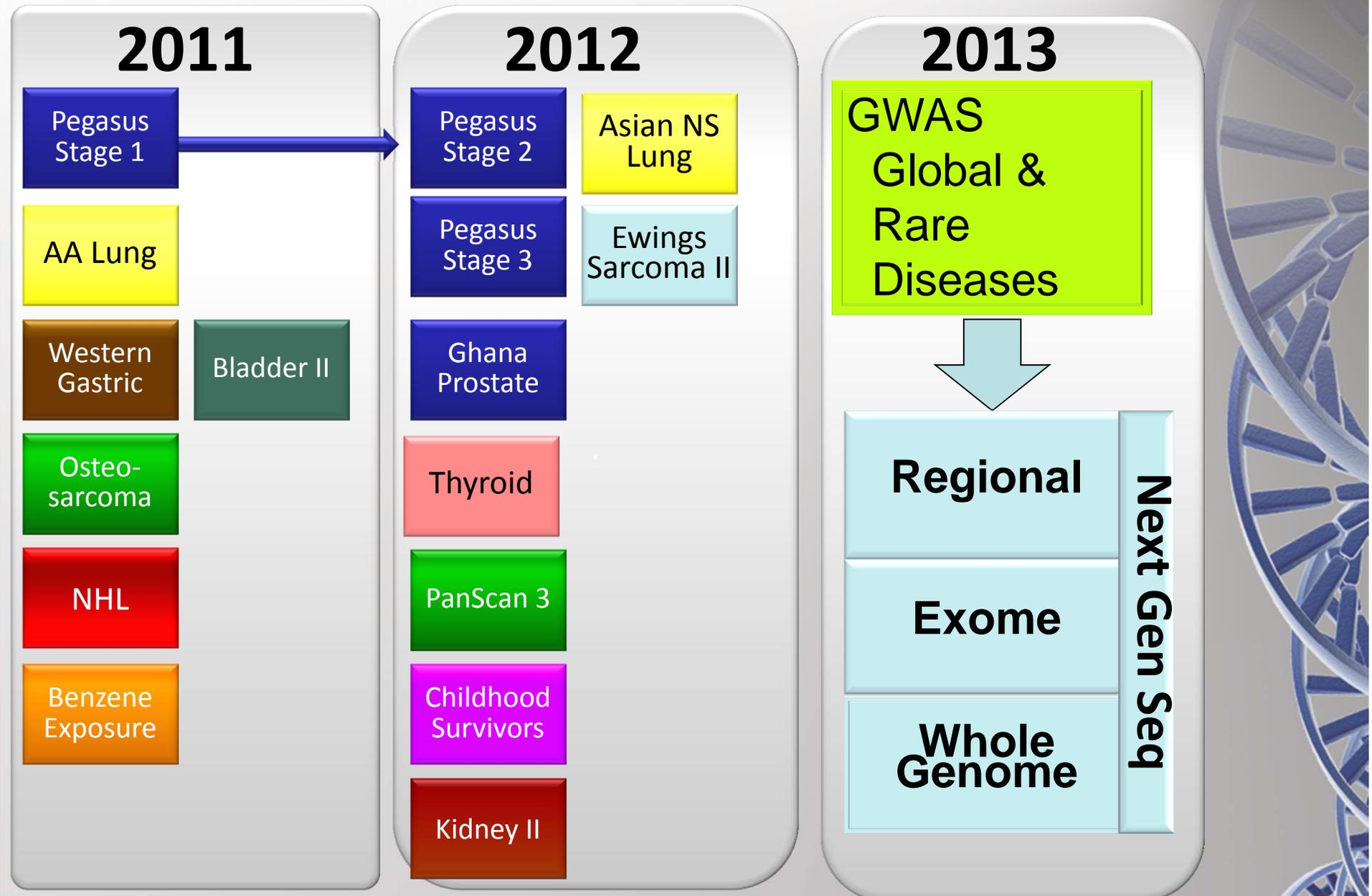
Analyzed & Delivered Data

SNP/CNV Genotypes:	76 x 10 ¹²
Regional Sequences:	100 Gbps
High-coverage exomes:	231, 2 Tbps aligned sequence, 200x avg coverage for Illumina HiSeq + Nimblegen 10-12x for Roche/454
Whole-genomes:	78, 15 Tbps aligned sequence, 60x avg coverage, Complete Genomics

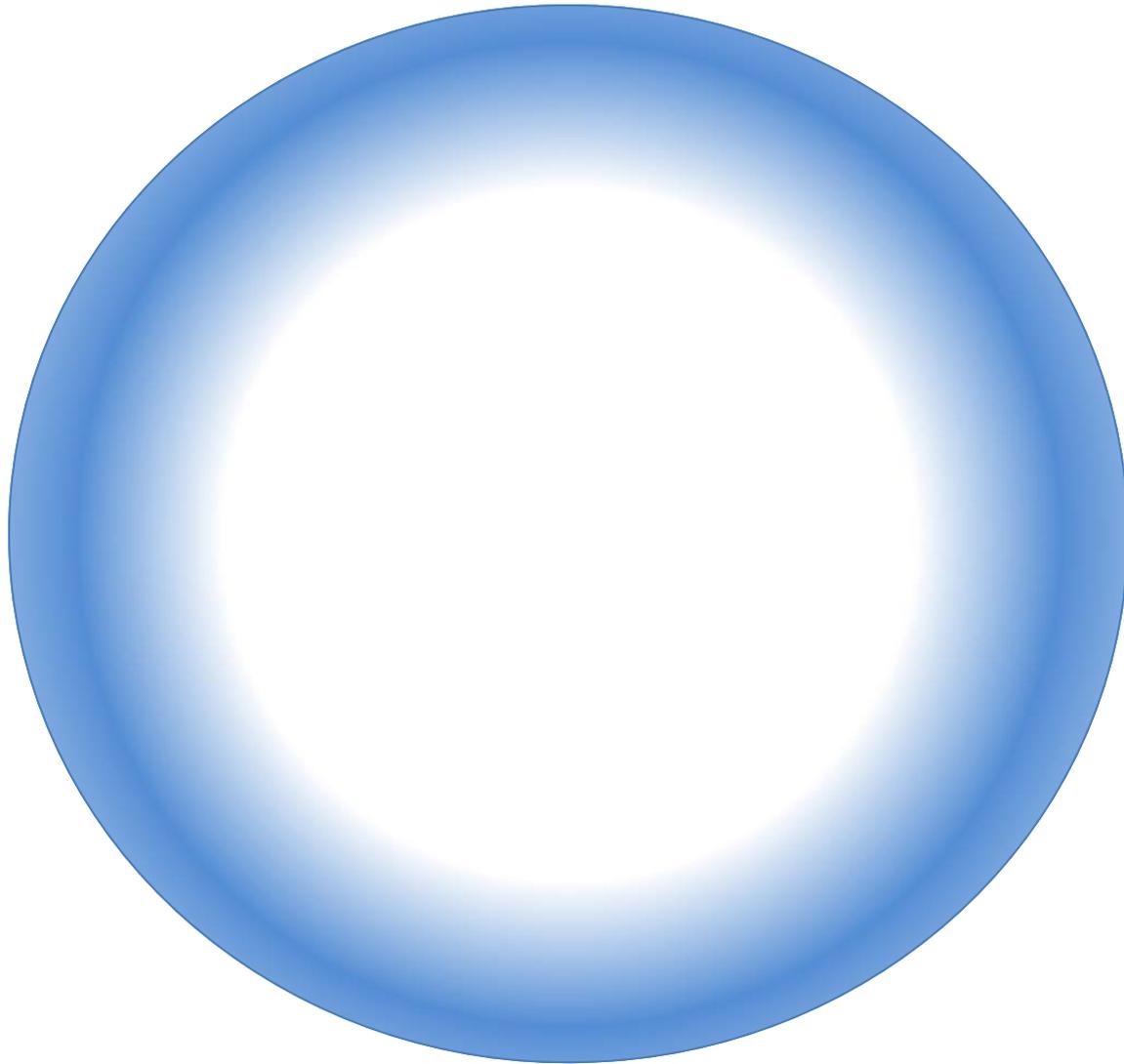
GWAS Timeline



GWAS->Sequencing Timeline



DCEG Total GWAS Set (TGS)



Resource based on DCEG 'TGS'

Zhaoming Wang, Kevin B Jacobs
Meredith Yeager, Amy Hutchinson
Joshua Sampson, Nilanjan Chatterjee,
Demetrius Albanes, Sonja I Berndt
Charles C Chung, W Ryan Diver
Susan M Gapstur, Lauren R Teras
Christopher A Haiman, Brian E Henderson,
Daniel Stram, Xiang Deng, Ann W Hsing,
Jarmo Virtamo, Michael A Eberle,
Jennifer L Stone, Mark P Purdue,
Phil Taylor, Margaret Tucker,
Stephen J Chanock

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Improved imputation of common and uncommon SNPs with a new reference set

Statistical imputation of genotype data is an important statistical technique that uses patterns of linkage disequilibrium observed in a reference set of haplotypes to computationally predict genetic variants *in silico*¹. Currently, the most popular reference sets are the publicly available International HapMap² and 1000 Genomes data sets³. Although these resources are valuable for imputing a sizeable fraction of common SNPs, they may not be optimal for imputing data for the next generation of genome-wide association studies (GWAS) and SNP arrays, which explore a fraction of uncommon variants.

We have built a new resource for the imputation of SNPs for existing and future GWAS, known as the Division of Cancer Epidemiology and Genetics (DCEG) Reference Set. The data set has genotypes for cancer-free individuals, including 728 of European ancestry from three large prospectively sampled studies^{4–6}, 98 African-American individuals from the Prostate, Lung, Colon and Ovary Cancer Screening Trial (PLCO), 74 Chinese individuals from a clinical trial in Shanxi, China (SHNX)⁷ and 349 individuals from the HapMap Project (Table 1). The final harmonized data set includes 2.8 million autosomal polymorphic SNPs for 1,249 individuals after rigorous quality control metrics were applied (see Supplementary Methods and Supplementary Tables 1 and 2).

We compared the imputation performance of the DCEG Reference Set to that of the International HapMap and 1000 Genomes reference sets, which are available from the IMPUTE2 website (see URLs). We assessed imputation accuracy by taking directly genotyped SNP data from the DCEG Reference Set and masking subsets to simulate data from two low-cost commercial genotyping arrays commonly used in GWAS studies (Illumina Human Hap660 and Human OmniExpress). Probabilistic genotypes were imputed using both IMPUTE2 (ref. 8) and BEAGLE⁹ software and compared with the masked genotyped SNPs. Accuracy was measured using the squared Pearson correlation coefficient (R^2) under an allelic dosage model (see Supplementary Methods). Using the new reference set, we observed higher imputation accuracy than that achieved with the

combination of 1000 Genomes and HapMap data across a spectrum of minor allele frequencies (MAFs) (Fig. 1). Accuracy in individuals of European ancestry imputed from Hap660 or OmniExpress arrays, measured by the proportion of variants imputed with $R^2 > 0.8$, improved by 34%, 23% and 12% for variants with MAFs of 3%, 5% and 10%, respectively. We estimated the difference in power to detect associations in GWAS designs between an imputed data set and one composed of directly genotyped SNPs with the DCEG Reference Set by adapting a model developed by Park et al.¹⁰. When using Hap660 data for imputation, we observed detection rates of 92.9% when imputing with the DCEG Reference Set and 84.7% with the 1000 Genomes and HapMap reference sets relative to the detection rate attained with directly genotyped SNPs; for OmniExpress data, we observed detection rates of 93.9% and 86.2% for these reference sets, respectively.

Because imputation accuracy depends on the similarity of haplotypes between

reference and study populations, we examined an extreme scenario in which we used a reference population from Finland (Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study, ATBC) to impute genotypes using OmniExpress data from a US population of European ancestry (PLCO) (Supplementary Fig. 1). For common SNPs, there was minimal loss of imputation accuracy when using the reference population from Finland relative to the US-based Cancer Prevention Study II (CPSII) or a combined population of HapMap individuals from Utah of Northern and Western European ancestry (CEU) and from northern Italy (Toscani in Italy, TSI). This result suggests that, for common variants, a reference set of sufficient size can adequately predict common SNPs when there is a discrepancy in population ancestry, provided that comparable haplotypes are sufficiently represented. This observation should enable investigators to proceed more confidently with imputation without additional genotyping in related but not identical populations.

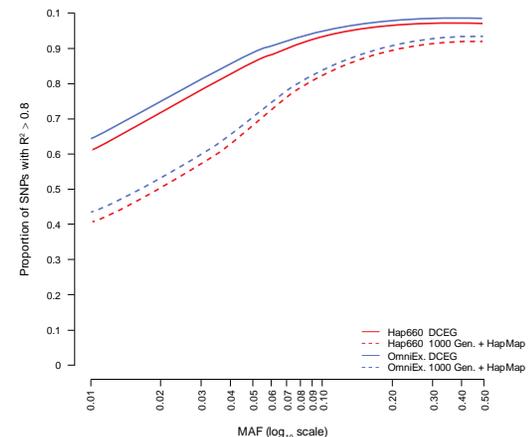


Figure 1 Imputation accuracy for individuals of European ancestry with the DCEG Reference Set and publicly available reference sets. The proportion of SNPs with allelic dosage $R^2 > 0.8$ by MAF is shown on the log scale to emphasize differences at smaller values. Red lines show imputation of Hap660 data, and blue lines show imputation of OmniExpress data. Solid lines, imputation using the DCEG Reference Set; dashed lines, imputation using the 1000 Genomes plus HapMap 3 reference sets.

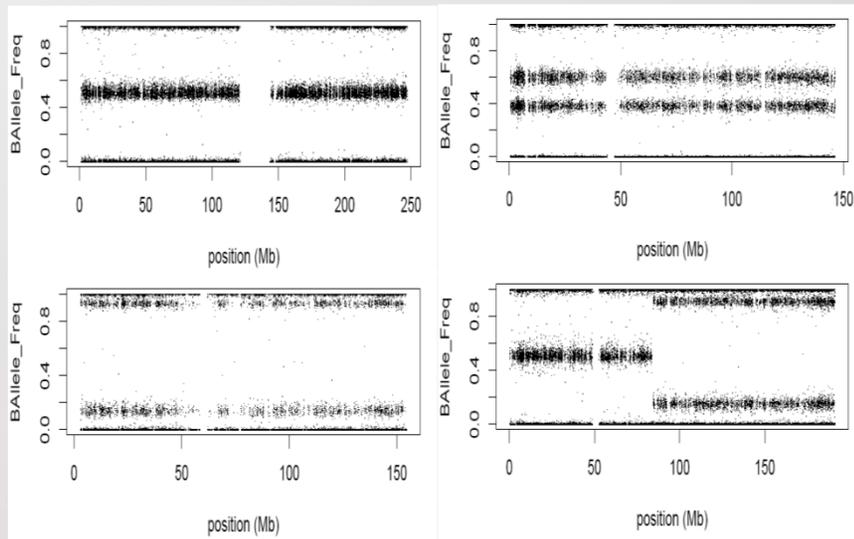
Unanticipated Directions



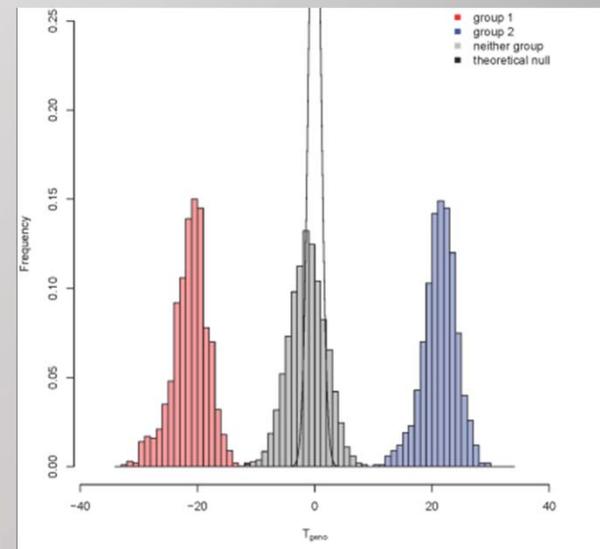
Genome-wide association studies

Large chromosomal abnormalities,
structural variation, aneuploidy in
Germ-line DNA

Privacy & Confidentiality
GWAS membership

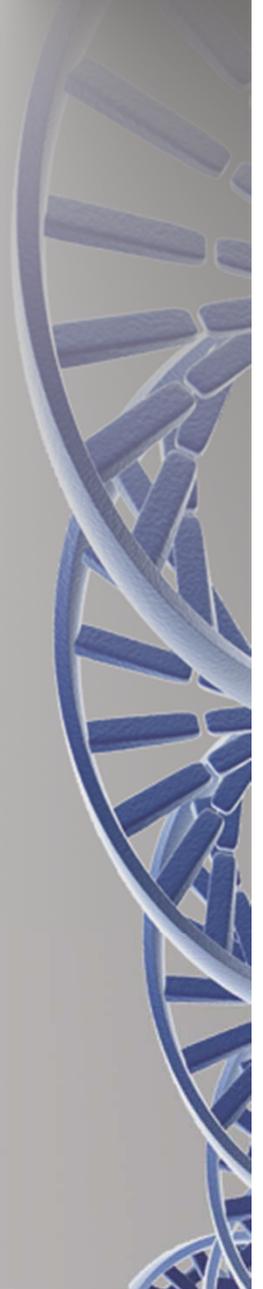
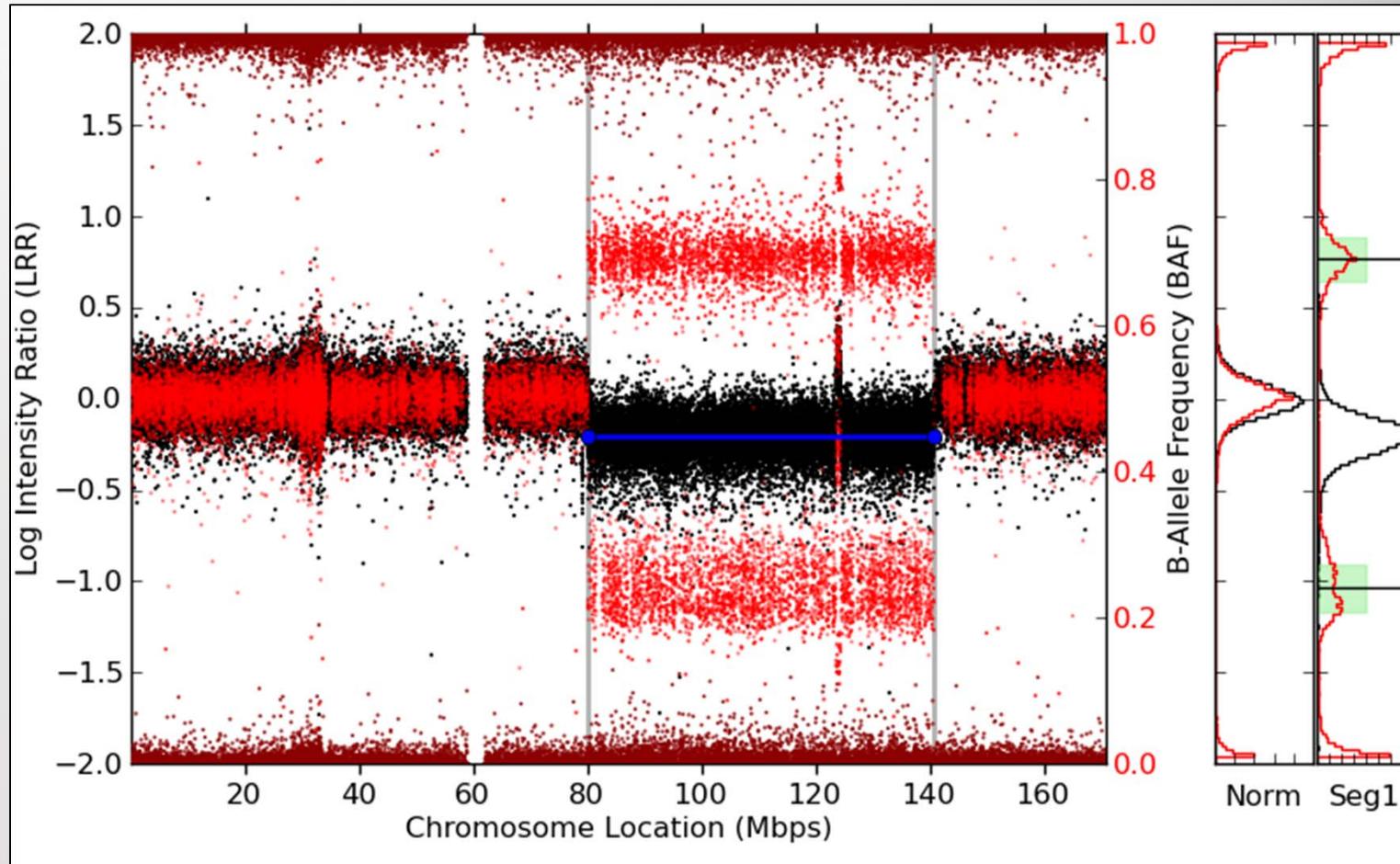


Rodriguez-Santiago AJHG 2010



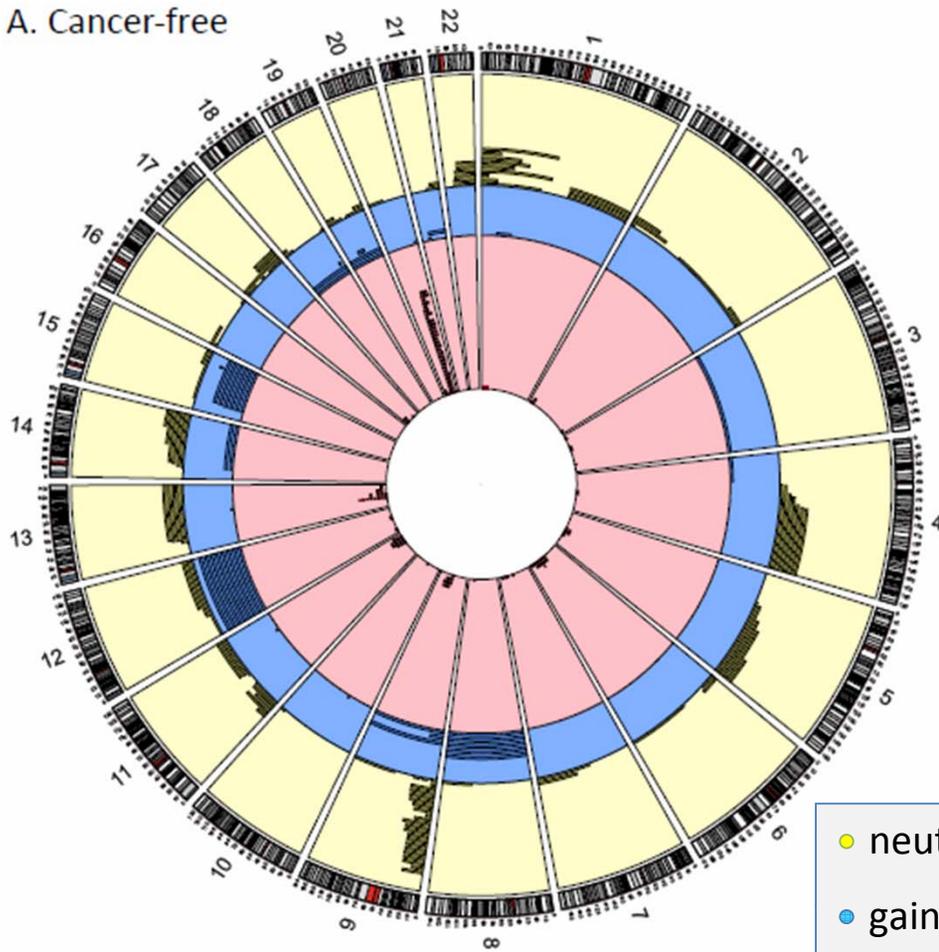
Jacobs Nature Genetics 2009

Mosaic Deletion

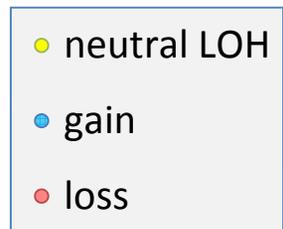
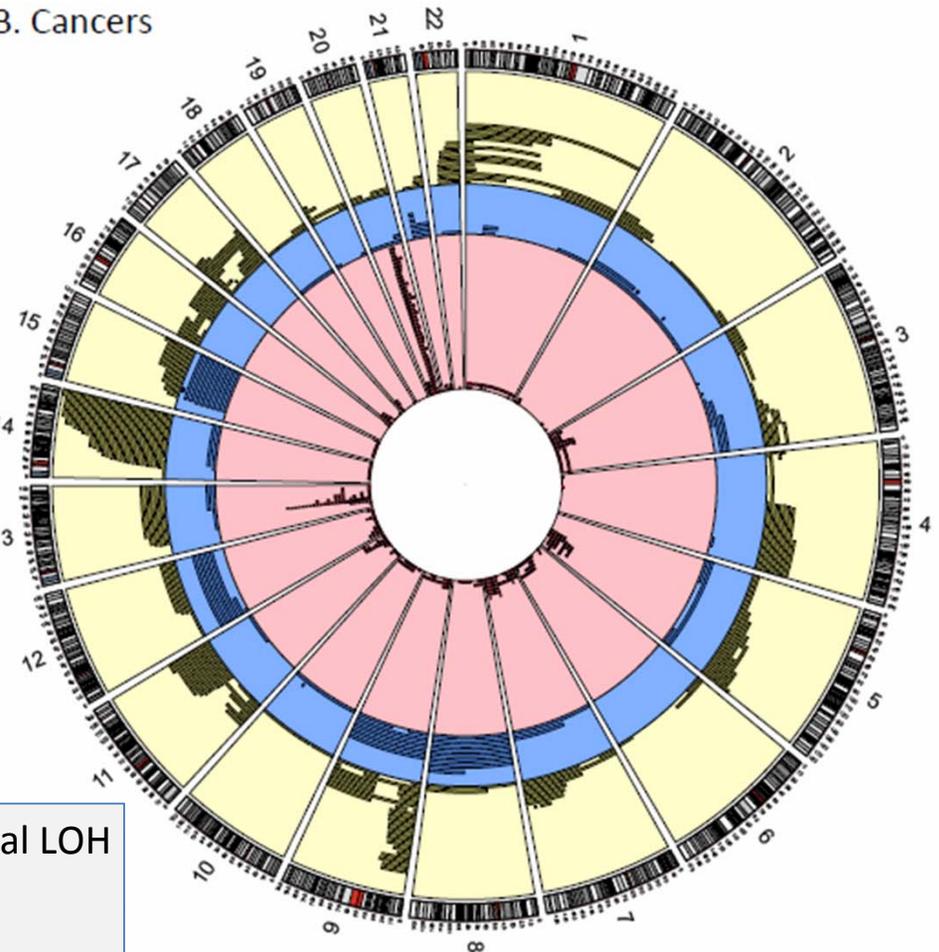


Circos Plot of large mosaic events (> 2 Mb) in 57,583 individuals

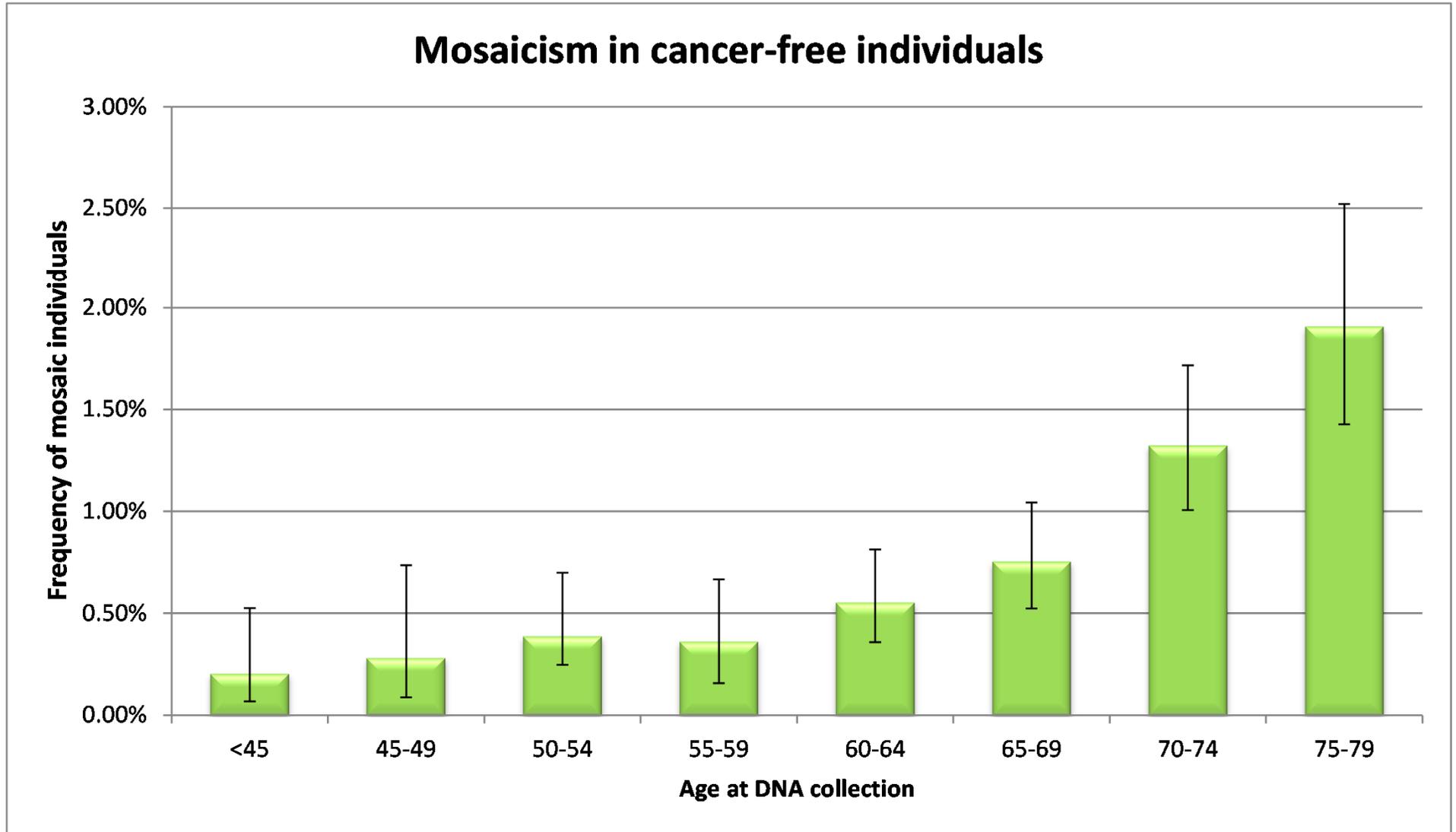
A. Cancer-free



B. Cancers



Age at DNA Collection is the Strongest Predictor of Genetic Mosaicism



CGF & Data Sharing



- Posted first public GWAS datasets for breast & prostate cancer
 - Aggregate data removed in 2008 in response to NIH policy change
- Led development of standards for GWAS posting with dbGaP
- Contributed all DCEG GWAS datasets to dbGaP
- CGF was instrumental in addressing privacy issues with GWAS and other high-dimensional aggregate genomics data

LETTERS

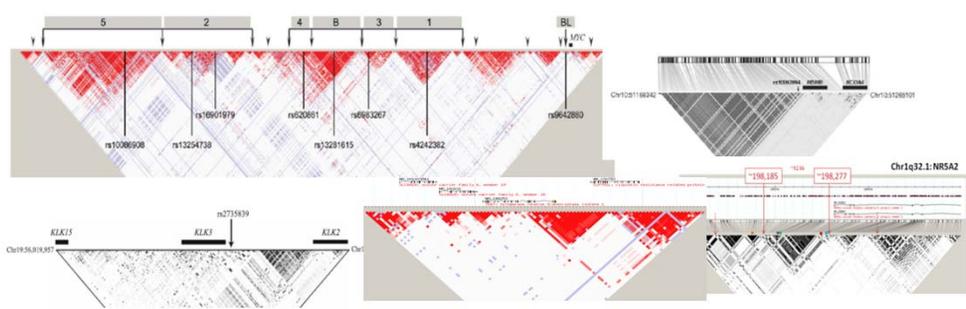
nature
genetics

A new statistic and its power to infer membership
in a genome-wide association study using genotype
frequencies

Kevin B Jacobs¹⁻³, Meredith Yeager^{1,2}, Sholom Wacholder², David Craig⁴, Peter Kraft⁵, David J Hunter⁵,
Justin Paschal⁶, Teri A Manolio⁷, Margaret Tucker², Robert N Hoover², Gilles D Thomas²,
Stephen J Chanock^{2,8} & Nilanjan Chatterjee^{2,8}

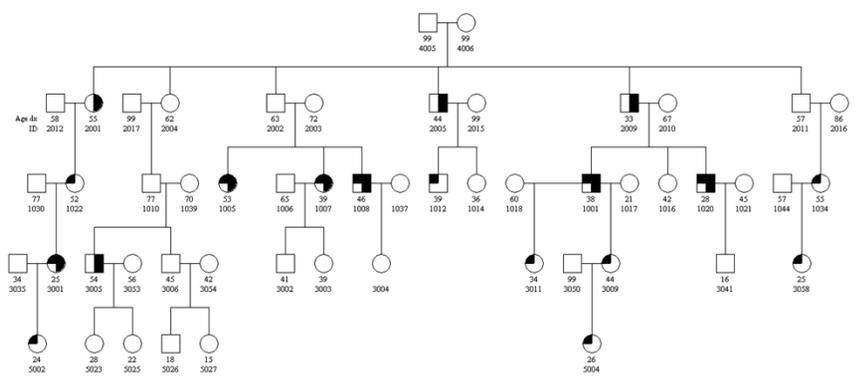
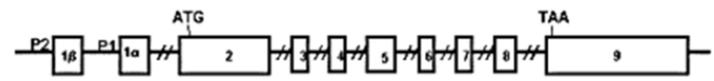
erved.

Regional GWAS and linkage follow-up

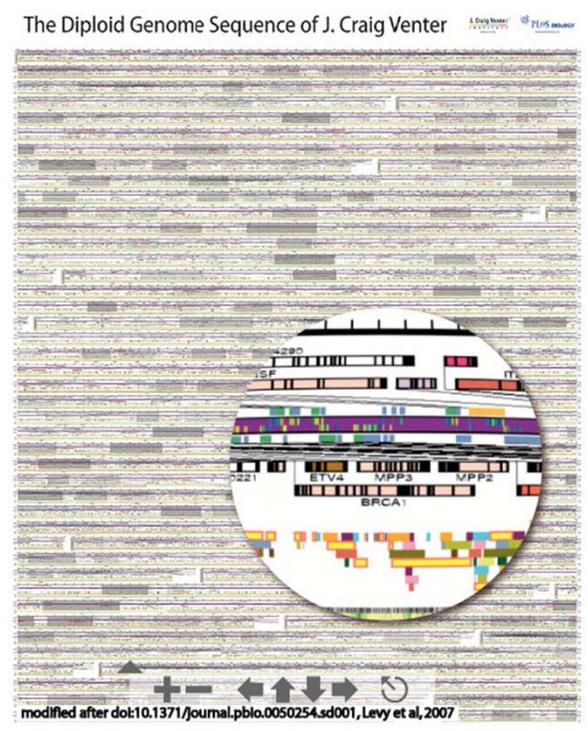


Candidate gene/exon

Sequencing



Whole exome



Whole genome

NGS Capabilities

Roche 454 GS FLX (2)

- Installed 2008
- Chosen for:
 - Read length
 - Multiplexing capability
- Current Output:
 - Multiplexing up to 264 samples
 - Average of 350-400bp/read



~~Illumina HiScan SQ~~

- ~~Installed August 2010~~



Illumina HiSeq 2000

- Installed April 2011
- Chosen for:
 - Throughput sufficient for exome/whole genome sequencing
- Current Output:
 - 300 Gbps/week (16 exomes)
 - 76-100 bp PE reads
- Expanded sequencing applications
 - CHiPseq
 - RNAseq

Life Technology/Ion Torrent PGM

- First Installed Jan 2011
- 6 machines as of Jan 2012
- Chosen for:
 - Cost
 - Reliability
 - Flexibility



Bumps along the way....

2007: Movement into ATP-SAIC

- Expectation of better alignment with program resources

2009: Movement out

- ATP Leadership sought to interrupt close collaboration and direct towards other business opportunities
- Placed under SAIC Research Administration OD

Recent Bump

- Sample handling bottleneck
 - CGF processes used for setting up DNA Extraction & Sample Handling Lab (DESL) in 2006
 - Increased demands stressed DESL
 - Stand alone service lab was realigned with CGF in 2011 due to
 - Quality Control Issues
 - Production Delays

Current Focus of Activities

Role of GWAS for:

1. Less common diseases w/ limited biospecimens
2. Complete our understanding of the contribution of common variant to cancer risk
 - Overall and population specific
3. Denser arrays for less common variation

Family & Special Population Analysis

- Exome & whole-genome sequencing
- Follow-up in families and unrelated subjects

Challenges Ahead

- Transition from GWAS to sequencing for investigation of germ-line susceptibility
- Further integration of environmental exposures
- Optimal storage, processing, and mining of whole-genome sequence data

Critical Mass

Analytical and Bioinformatic Expertise

- Close collaboration from inception to publication
 - Studies
 - Methodology
- Software development & dissemination
- Systematic data sharing
- Integrative analysis across studies & data types

Success of DCEG Core Genotyping Facility

- DCEG's decades of investment in epidemiology & genetics
 - Close collaborations between DCEG & FFRDC (CGF) epidemiologists, biostatisticians, geneticists, bioinformaticians and laboratory experts
- Dedicated facility framework